

## COURSE OUTLINES – BIOLOGY – SL + HL – 2023/2024 (DP1)

Level of organisation	Theme	Topic	Content	Allocated time		
				IB hours (60 minutes)	SL – 4 lessons per week	HL – 6 lessons per week
1. Molecules	A. Unity and Diversity	A1.1 Water	A1.1.1—Water as the medium for life A1.1.2—Hydrogen bonds as a consequence of the polar covalent bonds within water molecules A1.1.3—Cohesion of water molecules due to hydrogen bonding and consequences for organisms A1.1.4—Adhesion of water to materials that are polar or charged and impacts for organisms A1.1.5—Solvent properties of water linked to its role as a medium for metabolism and for transport in plants and animals A1.1.6—Physical properties of water and the consequences for animals in aquatic habitats A1.1.7—Extraterrestrial origin of water on Earth and reasons for its retention A1.1.8—Relationship between the search for extraterrestrial life and the presence of water	2+1	3	4

		A1.2 Nucleic acids	<p>A1.2.1—DNA as the genetic material of all living organisms</p> <p>A1.2.2—Components of a nucleotide</p> <p>A1.2.3—Sugar–phosphate bonding and the sugar–phosphate “backbone” of DNA and RNA</p> <p>A1.2.4—Bases in each nucleic acid that form the basis of a code</p> <p>A1.2.5—RNA as a polymer formed by condensation of nucleotide monomers</p> <p>A1.2.6—DNA as a double helix made of two antiparallel strands of nucleotides with two strands linked by hydrogen bonding between complementary base pairs</p> <p>A1.2.7—Differences between DNA and RNA</p> <p>A1.2.8—Role of complementary base pairing in allowing genetic information to be replicated and expressed</p> <p>A1.2.9—Diversity of possible DNA base sequences and the limitless capacity of DNA for storing information</p> <p>A1.2.10—Conservation of the genetic code across all life forms as evidence of universal common ancestry</p> <p>A1.2.11—Directionality of RNA and DNA</p> <p>A1.2.12—Purine-to-pyrimidine bonding as a component of DNA helix stability</p> <p>A1.2.13—Structure of a nucleosome</p> <p>A1.2.14—Evidence from the Hershey–Chase experiment for DNA as the genetic material</p> <p>A1.2.15—Chargaff’s data on the relative amounts of pyrimidine and purine bases across diverse life forms</p>	3+2	4	7
	<b>B. Form and function</b>	B1.1 Carbohydrates and lipids	B1.1.1—Chemical properties of a carbon atom allowing for the formation	4	5	5

			<p>of diverse compounds upon which life is based</p> <p>B1.1.2—Production of macromolecules by condensation reactions that link monomers to form a polymer</p> <p>B1.1.3—Digestion of polymers into monomers by hydrolysis reactions</p> <p>B1.1.4—Form and function of monosaccharides</p> <p>B1.1.5—Polysaccharides as energy storage compounds</p> <p>B1.1.6—Structure of cellulose related to its function as a structural polysaccharide in plants</p> <p>B1.1.7—Role of glycoproteins in cell–cell recognition</p> <p>B1.1.8—Hydrophobic properties of lipids</p> <p>B1.1.9—Formation of triglycerides and phospholipids by condensation reactions</p> <p>B1.1.10—Difference between saturated, monounsaturated and polyunsaturated fatty acids</p> <p>B1.1.11—Triglycerides in adipose tissues for energy storage and thermal insulation</p> <p>B1.1.12—Formation of phospholipid bilayers as a consequence of the hydrophobic and hydrophilic regions</p> <p>B1.1.13—Ability of non-polar steroids to pass through the phospholipid bilayer</p>			
		B1.2 Proteins	<p>B1.2.1—Generalized structure of an amino acid</p> <p>B1.2.2—Condensation reactions forming dipeptides and longer chains of amino acids</p> <p>B1.2.3—Dietary requirements for amino acids</p> <p>B1.2.4—Infinite variety of possible peptide chains</p>	2+2	3	6

			<p>B1.2.5—Effect of pH and temperature on protein structure</p> <p>B1.2.6—Chemical diversity in the R-groups of amino acids as a basis for the immense diversity in protein form and function</p> <p>B1.2.7—Impact of primary structure on the conformation of proteins</p> <p>B1.2.8—Pleating and coiling of secondary structure of proteins</p> <p>B1.2.9—Dependence of tertiary structure on hydrogen bonds, ionic bonds, disulfide covalent bonds and hydrophobic interactions</p> <p>B1.2.10—Effect of polar and non-polar amino acids on tertiary structure of proteins</p> <p>B1.2.11—Quaternary structure of non-conjugated and conjugated proteins</p> <p>B1.2.12—Relationship of form and function in globular and fibrous proteins</p>			
	<b>C. Interaction and interdependence</b>	C1.1 Enzymes and metabolism	<p>C1.1.1—Enzymes as catalysts</p> <p>C1.1.2—Role of enzymes in metabolism</p> <p>C1.1.3—Anabolic and catabolic reactions</p> <p>C1.1.4—Enzymes as globular proteins with an active site for catalysis</p> <p>C1.1.5—Interactions between substrate and active site to allow induced-fit binding</p> <p>C1.1.6—Role of molecular motion and substrate-active site collisions in enzyme catalysis</p> <p>C1.1.7—Relationships between the structure of the active site, enzyme–substrate specificity and denaturation</p> <p>C1.1.8—Effects of temperature, pH and substrate concentration on the rate of enzyme activity</p>	3+2	4	7

			<p>C1.1.9—Measurements in enzyme-catalysed reactions</p> <p>C1.1.10—Effect of enzymes on activation energy</p> <p>C1.1.11—Intracellular and extracellular enzyme-catalysed reactions</p> <p>C1.1.12—Generation of heat energy by the reactions of metabolism</p> <p>C1.1.13—Cyclical and linear pathways in metabolism</p> <p>C1.1.14—Allosteric sites and non-competitive inhibition</p> <p>C1.1.15—Competitive inhibition as a consequence of an inhibitor binding reversibly to an active site</p> <p>C1.1.16—Regulation of metabolic pathways by feedback inhibition</p> <p>C1.1.17—Mechanism-based inhibition as a consequence of chemical changes to the active site caused by the irreversible binding of an inhibitor</p>			
		C1.2 Cell respiration	<p>C1.2.1—ATP as the molecule that distributes energy within cells</p> <p>C1.2.2—Life processes within cells that ATP supplies with energy</p> <p>C1.2.3—Energy transfers during interconversions between ATP and ADP</p> <p>C1.2.4—Cell respiration as a system for producing ATP within the cell using energy released from carbon compounds</p> <p>C1.2.5—Differences between anaerobic and aerobic cell respiration in humans</p> <p>C1.2.6—Variables affecting the rate of cell respiration</p> <p>C1.2.7—Role of NAD as a carrier of hydrogen and oxidation by removal of hydrogen during cell respiration</p> <p>C1.2.8—Conversion of glucose to pyruvate by stepwise reactions in glycolysis with a net yield of ATP and reduced NAD</p>	2+3	3	7

			<p>C1.2.9—Conversion of pyruvate to lactate as a means of regenerating NAD in anaerobic cell respiration</p> <p>C1.2.10—Anaerobic cell respiration in yeast and its use in brewing and baking</p> <p>C1.2.11—Oxidation and decarboxylation of pyruvate as a link reaction in aerobic cell respiration</p> <p>C1.2.12—Oxidation and decarboxylation of acetyl groups in the Krebs cycle with a yield of ATP and reduced NAD</p> <p>C1.2.13—Transfer of energy by reduced NAD to the electron transport chain in the mitochondrion</p> <p>C1.2.14—Generation of a proton gradient by flow of electrons along the electron transport chain</p> <p>C1.2.15—Chemiosmosis and the synthesis of ATP in the mitochondrion</p> <p>C1.2.16—Role of oxygen as terminal electron acceptor in aerobic cell respiration</p> <p>C.1.2.17—Differences between lipids and carbohydrates as respiratory substrates</p>			
		C1.3 Photosynthesis	<p>C1.3.1—Transformation of light energy to chemical energy when carbon compounds are produced in photosynthesis</p> <p>C1.3.2—Conversion of carbon dioxide to glucose in photosynthesis using hydrogen obtained by splitting water</p> <p>C1.3.3—Oxygen as a by-product of photosynthesis in plants, algae and cyanobacteria</p> <p>C1.3.4—Separation and identification of photosynthetic pigments by chromatography</p>	3+3	4	8

			<p>C1.3.5—Absorption of specific wavelengths of light by photosynthetic pigments</p> <p>C1.3.6—Similarities and differences of absorption and action spectra</p> <p>C1.3.7—Techniques for varying concentrations of carbon dioxide, light intensity or temperature experimentally to investigate the effects of limiting factors on the rate of photosynthesis</p> <p>C1.3.8—Carbon dioxide enrichment experiments as a means of predicting future rates of photosynthesis and plant growth</p> <p>C1.3.9—Photosystems as arrays of pigment molecules that can generate and emit excited electrons</p> <p>C1.3.10—Advantages of the structured array of different types of pigment molecules in a photosystem</p> <p>C1.3.11—Generation of oxygen by the photolysis of water in photosystem II</p> <p>C1.3.12—ATP production by chemiosmosis in thylakoids</p> <p>C1.3.13—Reduction of NADP by photosystem I</p> <p>C1.3.14—Thylakoids as systems for performing the light-dependent reactions of photosynthesis</p> <p>C1.3.15—Carbon fixation by Rubisco</p> <p>C1.3.16—Synthesis of triose phosphate using reduced NADP and ATP</p> <p>C1.3.17—Regeneration of RuBP in the Calvin cycle using ATP</p> <p>C1.3.18—Synthesis of carbohydrates, amino acids and other carbon compounds using the products of the Calvin cycle and mineral nutrients</p> <p>C1.3.19—Interdependence of the light-dependent and light-independent reactions</p>			
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	<b>D. Continuity and change</b>	D1.1 DNA replication	<p>D1.1.1—DNA replication as production of exact copies of DNA with identical base sequences</p> <p>D1.1.2—Semi-conservative nature of DNA replication and role of complementary base pairing</p> <p>D1.1.3—Role of helicase and DNA polymerase in DNA replication</p> <p>D1.1.4—Polymerase chain reaction and gel electrophoresis as tools for amplifying and separating DNA</p> <p>D1.1.5—Applications of polymerase chain reaction and gel electrophoresis</p> <p>D1.1.6—Directionality of DNA polymerases</p> <p>D1.1.7—Differences between replication on the leading strand and the lagging strand</p> <p>D1.1.8—Functions of DNA primase, DNA polymerase I, DNA polymerase III and DNA ligase in replication</p> <p>D1.1.9—DNA proofreading</p>	2+2	3	6
		D1.2 Protein synthesis	<p>D1.2.1—Transcription as the synthesis of RNA using a DNA template</p> <p>D1.2.2—Role of hydrogen bonding and complementary base pairing in transcription</p> <p>D1.2.3—Stability of DNA templates</p> <p>D1.2.4—Transcription as a process required for the expression of genes</p> <p>D1.2.5—Translation as the synthesis of polypeptides from mRNA</p> <p>D1.2.6—Roles of mRNA, ribosomes and tRNA in translation</p> <p>D1.2.7—Complementary base pairing between tRNA and mRNA</p> <p>D1.2.8—Features of the genetic code</p> <p>D1.2.9—Using the genetic code expressed as a table of mRNA codons</p> <p>D1.2.10—Stepwise movement of the ribosome along mRNA and linkage of</p>	3+3	4	8

			<p>amino acids by peptide bonding to the growing polypeptide chain</p> <p>D1.2.11—Mutations that change protein structure</p> <p>D1.2.12—Directionality of transcription and translation</p> <p>D1.2.13—Initiation of transcription at the promoter</p> <p>D1.2.14—Non-coding sequences in DNA do not code for polypeptides</p> <p>D1.2.15—Post-transcriptional modification in eukaryotic cells</p> <p>D1.2.16—Alternative splicing of exons to produce variants of a protein from a single gene</p> <p>D1.2.17—Initiation of translation</p> <p>D1.2.18—Modification of polypeptides into their functional state</p> <p>D1.2.19—Recycling of amino acids by proteasomes</p>			
		D1.3 Mutations and gene editing	<p>D1.3.1—Gene mutations as structural changes to genes at the molecular level</p> <p>D1.3.2—Consequences of base substitutions</p> <p>D1.3.3—Consequences of insertions and deletions</p> <p>D1.3.4—Causes of gene mutation</p> <p>D1.3.5—Randomness in mutation</p> <p>D1.3.6—Consequences of mutation in germ cells and somatic cells</p> <p>D1.3.7—Mutation as a source of genetic variation</p> <p>D1.3.8—Gene knockout as a technique for investigating the function of a gene by changing it to make it inoperative</p> <p>D1.3.9—Use of the CRISPR sequences and the enzyme Cas9 in gene editing</p> <p>D1.3.10—Hypotheses to account for conserved or highly conserved sequences in genes</p>	3+2	4	7

Practical work				12+12	16	31
2. Cells	A. Unity and Diversity	A2.1 Origins of cells (HL only)	<p>A2.1.1—Conditions on early Earth and the pre-biotic formation of carbon compounds</p> <p>A2.1.2—Cells as the smallest units of self-sustaining life</p> <p>A2.1.3—Challenge of explaining the spontaneous origin of cells</p> <p>A2.1.4—Evidence for the origin of carbon compounds</p> <p>A2.1.5—Spontaneous formation of vesicles by coalescence of fatty acids into spherical bilayers</p> <p>A2.1.6—RNA as a presumed first genetic material</p> <p>A2.1.7—Evidence for a last universal common ancestor</p> <p>A2.1.8—Approaches used to estimate dates of the first living cells and the last universal common ancestor</p> <p>A2.1.9—Evidence for the evolution of the last universal common ancestor in the vicinity of hydrothermal vents</p>	2		3
		A2.2 Cell structure	<p>A2.2.1—Cells as the basic structural unit of all living organisms</p> <p>A2.2.2—Microscopy skills</p> <p>A2.2.3—Developments in microscopy</p> <p>A2.2.4—Structures common to cells in all living organisms</p> <p>A2.2.5—Prokaryote cell structure</p> <p>A2.2.6—Eukaryote cell structure</p> <p>A2.2.7—Processes of life in unicellular organisms</p> <p>A2.2.8—Differences in eukaryotic cell structure between animals, fungi and plants</p> <p>A2.2.9—Atypical cell structure in eukaryotes</p>	4+1	5	7

			<p>A2.2.10—Cell types and cell structures viewed in light and electron micrographs</p> <p>A2.2.11—Drawing and annotation based on electron micrographs</p> <p>A2.2.12—Origin of eukaryotic cells by endosymbiosis</p> <p>A2.2.13—Cell differentiation as the process for developing specialized tissues in multicellular organisms</p> <p>A2.2.14—Evolution of multicellularity</p>			
		A2.3 Viruses (HL only)	<p>A2.3.1—Structural features common to viruses</p> <p>A2.3.2—Diversity of structure in viruses</p> <p>A2.3.3—Lytic cycle of a virus</p> <p>A2.3.4—Lysogenic cycle of a virus</p> <p>A2.3.5—Evidence for several origins of viruses from other organisms</p> <p>A2.3.6—Rapid evolution in viruses</p>	2		3
	<b>B. Form and function</b>	B2.1 Membranes and membrane transport	<p>B2.1.1—Lipid bilayers as the basis of cell membranes</p> <p>B2.1.2—Lipid bilayers as barriers</p> <p>B2.1.3—Simple diffusion across membranes</p> <p>B2.1.4—Integral and peripheral proteins in membranes</p> <p>B2.1.5—Movement of water molecules across membranes by osmosis and the role of aquaporins</p> <p>B2.1.6—Channel proteins for facilitated diffusion</p> <p>B2.1.7—Pump proteins for active transport</p> <p>B2.1.8—Selectivity in membrane permeability</p> <p>B2.1.9—Structure and function of glycoproteins and glycolipids</p> <p>B2.1.10—Fluid mosaic model of membrane structure</p>	4+2	5	8

			<p>B2.1.11—Relationships between fatty acid composition of lipid bilayers and their fluidity</p> <p>B2.1.12—Cholesterol and membrane fluidity in animal cells</p> <p>B2.1.13—Membrane fluidity and the fusion and formation of vesicles</p> <p>B2.1.14—Gated ion channels in neurons</p> <p>B2.1.15—Sodium–potassium pumps as an example of exchange transporters</p> <p>B2.1.16—Sodium-dependent glucose cotransporters as an example of indirect active transport</p> <p>B2.1.17—Adhesion of cells to form tissues</p>			
		B2.2 Organelles and compartmentalization	<p>B2.2.1—Organelles as discrete subunits of cells that are adapted to perform specific functions</p> <p>B2.2.2—Advantage of the separation of the nucleus and cytoplasm into separate compartments</p> <p>B2.2.3—Advantages of compartmentalization in the cytoplasm of cells</p> <p>B2.2.4—Adaptations of the mitochondrion for production of ATP by aerobic cell respiration</p> <p>B2.2.5—Adaptations of the chloroplast for photosynthesis</p> <p>B2.2.6—Functional benefits of the double membrane of the nucleus</p> <p>B2.2.7—Structure and function of free ribosomes and of the rough endoplasmic reticulum</p> <p>B2.2.8—Structure and function of the Golgi apparatus</p> <p>B2.2.9—Structure and function of vesicles in cells</p>	1+2	1	3
		B2.3 Cell specialization	B2.3.1—Production of unspecialized cells following fertilization and their	2+1	3	4

			<p>development into specialized cells by differentiation</p> <p>B2.3.2—Properties of stem cells</p> <p>B2.3.3—Location and function of stem cell niches in adult humans</p> <p>B2.3.4—Differences between totipotent, pluripotent and multipotent stem cells</p> <p>B2.3.5—Cell size as an aspect of specialization</p> <p>B2.3.6—Surface area-to-volume ratios and constraints on cell size</p> <p>B2.3.7—Adaptations to increase surface area-to-volume ratios of cells</p> <p>B2.3.8—Adaptations of type I and type II pneumocytes in alveoli</p> <p>B2.3.9—Adaptations of cardiac muscle cells and striated muscle fibres</p> <p>B2.3.10—Adaptations of sperm and egg cells</p>			
	<b>C. Interaction and interdependence</b>	C2.1 Chemical signalling (HL only)	<p>C2.1.1—Receptors as proteins with binding sites for specific signalling chemicals</p> <p>C.2.1.2—Cell signalling by bacteria in quorum sensing</p> <p>C2.1.3—Hormones, neurotransmitters, cytokines and calcium ions as examples of functional categories of signalling chemicals in animals</p> <p>C2.1.4—Chemical diversity of hormones and neurotransmitters</p> <p>C2.1.5—Localized and distant effects of signalling molecules</p> <p>C2.1.6—Differences between transmembrane receptors in a plasma membrane and intracellular receptors in the cytoplasm or nucleus</p> <p>C2.1.7—Initiation of signal transduction pathways by receptors</p> <p>C2.1.8—Transmembrane receptors for neurotransmitters and changes to membrane potential</p>	4		5

			<p>C2.1.9—Transmembrane receptors that activate G proteins</p> <p>C2.1.10—Mechanism of action of epinephrine (adrenaline) receptors</p> <p>C2.1.11—Transmembrane receptors with tyrosine kinase activity</p> <p>C2.1.12—Intracellular receptors that affect gene expression</p> <p>C2.1.13—Effects of the hormones oestradiol and progesterone on target cells</p> <p>C2.1.14—Regulation of cell signalling pathways by positive and negative feedback</p>			
		C2.2Neural signalling	<p>C2.2.1—Neurons as cells within the nervous system that carry electrical impulses</p> <p>C2.2.2—Generation of the resting potential by pumping to establish and maintain concentration gradients of sodium and potassium ions</p> <p>C2.2.3—Nerve impulses as action potentials that are propagated along nerve fibres</p> <p>C2.2.4—Variation in the speed of nerve impulses</p> <p>C2.2.5—Synapses as junctions between neurons and between neurons and effector cells</p> <p>C2.2.6—Release of neurotransmitters from a presynaptic membrane</p> <p>C2.2.7—Generation of an excitatory postsynaptic potential</p> <p>C2.2.8—Depolarization and repolarization during action potentials</p> <p>C2.2.9—Propagation of an action potential along a nerve fibre/axon as a result of local currents</p> <p>C2.2.10—Oscilloscope traces showing resting potentials and action potentials</p>	3+3	4	8

			<p>C2.2.11—Saltatory conduction in myelinated fibres to achieve faster impulses</p> <p>C2.2.12—Effects of exogenous chemicals on synaptic transmission</p> <p>C2.2.13—Inhibitory neurotransmitters and generation of inhibitory postsynaptic potentials</p> <p>C2.2.14—Summation of the effects of excitatory and inhibitory neurotransmitters in a postsynaptic neuron</p> <p>C2.2.15—Perception of pain by neurons with free nerve endings in the skin</p> <p>C2.2.16—Consciousness as a property that emerges from the interaction of individual neurons in the brain</p>			
	<b>D. Continuity and change</b>	D2.1 Cell and nuclear division	<p>D2.1.1—Generation of new cells in living organisms by cell division</p> <p>D2.1.2—Cytokinesis as splitting of cytoplasm in a parent cell between daughter cells</p> <p>D2.1.3—Equal and unequal cytokinesis</p> <p>D2.1.4—Roles of mitosis and meiosis in eukaryotes</p> <p>D2.1.5—DNA replication as a prerequisite for both mitosis and meiosis</p> <p>D2.1.6—Condensation and movement of chromosomes as shared features of mitosis and meiosis</p> <p>D2.1.7—Phases of mitosis</p> <p>D2.1.8—Identification of phases of mitosis</p> <p>D2.1.9—Meiosis as a reduction division</p> <p>D2.1.10—Down syndrome and non-disjunction</p> <p>D2.1.11—Meiosis as a source of variation</p> <p>D2.1.12—Cell proliferation for growth, cell replacement and tissue repair</p> <p>D2.1.13—Phases of the cell cycle</p>	3+1	4	7

			<p>D2.1.14—Cell growth during interphase</p> <p>D2.1.15—Control of the cell cycle using cyclins</p> <p>D2.1.16—Consequences of mutations in genes that control the cell cycle</p> <p>D2.1.17—Differences between tumours in rates of cell division and growth and in the capacity for metastasis and invasion of neighbouring tissue</p>			
		D2.2 Gene expression (HL only)	<p>D2.2.1—Gene expression as the mechanism by which information in genes has effects on the phenotype</p> <p>D2.2.2—Regulation of transcription by proteins that bind to specific base sequences in DNA</p> <p>D2.2.3—Control of the degradation of mRNA as a means of regulating translation</p> <p>D2.2.4—Epigenesis as the development of patterns of differentiation in the cells of a multicellular organism</p> <p>D2.2.5—Differences between the genome, transcriptome and proteome of individual cells</p> <p>D2.2.6—Methylation of the promoter and histones in nucleosomes as examples of epigenetic tags</p> <p>D2.2.7—Epigenetic inheritance through heritable changes to gene expression</p> <p>D2.2.8—Examples of environmental effects on gene expression in cells and organisms</p> <p>D2.2.9—Consequences of removal of most but not all epigenetic tags from the ovum and sperm</p> <p>D2.2.10—Monozygotic twin studies</p> <p>D2.2.11—External factors impacting the pattern of gene expression</p>	3		4
		D2.3 Water potential	D2.3.1—Solvation with water as the solvent	2+2	3	6

			<p>D2.3.2—Water movement from less concentrated to more concentrated solutions</p> <p>D2.3.3—Water movement by osmosis into or out of cells</p> <p>D2.3.4—Changes due to water movement in plant tissue bathed in hypotonic and those bathed in hypertonic solutions</p> <p>D2.3.5—Effects of water movement on cells that lack a cell wall</p> <p>D2.3.6—Effects of water movement on cells with a cell wall</p> <p>D2.3.7—Medical applications of isotonic solutions</p> <p>D2.3.8—Water potential as the potential energy of water per unit volume</p> <p>D2.3.9—Movement of water from higher to lower water potential</p> <p>D2.3.10—Contributions of solute potential and pressure potential to the water potential of cells with walls</p> <p>D2.3.11—Water potential and water movements in plant tissue</p>			
<b>Collaborative Sciences Project</b>				10	13	13
<b>3. Organisms</b>	<b>A. Unity and Diversity</b>	A3.1 Diversity of organisms	<p>A3.1.1—Variation between organisms as a defining feature of life</p> <p>A3.1.2—Species as groups of organisms with shared traits</p> <p>A3.1.3—Binomial system for naming organisms</p> <p>A3.1.4—Biological species concept</p> <p>A3.1.5—Difficulties distinguishing between populations and species due to divergence of non-interbreeding populations during speciation</p> <p>A3.1.6—Diversity in chromosome numbers of plant and animal species</p> <p>A3.1.7—Karyotyping and karyograms</p>	3+2	4	7

			<p>A3.1.8—Unity and diversity of genomes within species</p> <p>A3.1.9—Diversity of eukaryote genomes</p> <p>A3.1.10—Comparison of genome sizes</p> <p>A3.1.11—Current and potential future uses of whole genome sequencing</p> <p>A3.1.12—Difficulties applying the biological species concept to asexually reproducing species and to bacteria that have horizontal gene transfer</p> <p>A3.1.13—Chromosome number as a shared trait within a species</p> <p>A3.1.14—Engagement with local plant or animal species to develop a dichotomous key</p> <p>A3.1.15—Identification of species from environmental DNA in a habitat using barcodes</p>			
		A3.2 Classification and cladistics (HL only)	<p>A3.2.1—Need for classification of organisms</p> <p>A3.2.2—Difficulties classifying organisms into the traditional hierarchy of taxa</p> <p>A3.2.3—Advantages of classification corresponding to evolutionary relationships</p> <p>A3.2.4—Clades as groups of organisms with common ancestry and shared characteristics</p> <p>A3.2.5—Gradual accumulation of sequence differences as the basis for estimates of when clades diverged from a common ancestor</p> <p>A3.2.6—Base sequences of genes or amino acid sequences of proteins as the basis for constructing cladograms</p> <p>A3.2.7—Analysing cladograms</p> <p>A3.2.8—Using cladistics to investigate whether the classification of groups</p>	3		4

			<p>corresponds to evolutionary relationships</p> <p>A3.2.9—Classification of all organisms into three domains using evidence from rRNA base sequences</p>			
<b>B. Form and function</b>	B3.1 Gas exchange	<p>B3.1.1—Gas exchange as a vital function in all organisms</p> <p>B3.1.2—Properties of gas-exchange surfaces</p> <p>B3.1.3—Maintenance of concentration gradients at exchange surfaces in animals</p> <p>B3.1.4—Adaptations of mammalian lungs for gas exchange</p> <p>B3.1.5—Ventilation of the lungs</p> <p>B3.1.6—Measurement of lung volumes</p> <p>B3.1.7—Adaptations for gas exchange in leaves</p> <p>B3.1.8—Distribution of tissues in a leaf</p> <p>B3.1.9—Transpiration as a consequence of gas exchange in a leaf</p> <p>B3.1.10—Stomatal density</p> <p>B3.1.11—Adaptations of foetal and adult haemoglobin for the transport of oxygen</p> <p>B3.1.12—Bohr shift</p> <p>B3.1.13—Oxygen dissociation curves as a means of representing the affinity of haemoglobin for oxygen at different oxygen concentrations</p>	3+1	4	5	
	B3.2 Transport	<p>B3.2.1—Adaptations of capillaries for exchange of materials between blood and the internal or external environment</p> <p>B3.2.2—Structure of arteries and veins</p> <p>B3.2.3—Adaptations of arteries for the transport of blood away from the heart</p> <p>B3.2.4—Measurement of pulse rates</p> <p>B3.2.5—Adaptations of veins for the return of blood to the heart</p>	3+2	4	7	

			<p>B3.2.6—Causes and consequences of occlusion of the coronary arteries</p> <p>B3.2.7—Transport of water from roots to leaves during transpiration</p> <p>B3.2.8—Adaptations of xylem vessels for transport of water</p> <p>B3.2.9—Distribution of tissues in a transverse section of the stem of a dicotyledonous plant</p> <p>B3.2.10—Distribution of tissues in a transverse section of the root of a dicotyledonous plant</p> <p>B3.2.11—Release and reuptake of tissue fluid in capillaries</p> <p>B3.2.12—Exchange of substances between tissue fluid and cells in tissues</p> <p>B3.2.13—Drainage of excess tissue fluid into lymph ducts</p> <p>B3.2.14—Differences between the single circulation of bony fish and the double circulation of mammals</p> <p>B3.2.15—Adaptations of the mammalian heart for delivering pressurized blood to the arteries</p> <p>B3.2.16—Stages in the cardiac cycle</p> <p>B3.2.17—Generation of root pressure in xylem vessels by active transport of mineral ions</p> <p>B3.2.18—Adaptations of phloem sieve tubes and companion cells for translocation of sap</p>			
		B3.3Muscle and motility (HL only)	<p>B3.3.1—Adaptations for movement as a universal feature of living organisms</p> <p>B3.3.2—Sliding filament model of muscle contraction</p> <p>B3.3.3—Role of the protein titin and antagonistic muscles in muscle relaxation</p> <p>B3.3.4—Structure and function of motor units in skeletal muscle</p>	3		4

			<p>B3.3.5—Roles of skeletons as anchorage for muscles and as levers</p> <p>B3.3.6—Movement at a synovial joint</p> <p>B3.3.7—Range of motion of a joint</p> <p>B3.3.8—Internal and external intercostal muscles as an example of antagonistic muscle action to facilitate internal body movements</p> <p>B3.3.9—Reasons for locomotion</p> <p>B3.3.10—Adaptations for swimming in marine mammals</p>			
<b>Total</b>				<b>76 hours SL/ 140 hours HL</b>	<b>101 lessons</b>	<b>186 lessons</b>

**COURSE OUTLINES – BIOLOGY – SL + HL – 2024/2025 (DP2)**

Level of organisation	Theme	Topic	Content	Allocated time		
				IB hours (60 minutes)	SL – 4 lessons per week	HL – 4 lessons per week
3. Organisms	C. Interaction and interdependence	C3.1 Integration of body systems	C3.1.1—System integration C3.1.2—Cells, tissues, organs and body systems as a hierarchy of subsystems that are integrated in a multicellular living organism C3.1.3—Integration of organs in animal bodies by hormonal and nervous signalling and by transport of materials and energy C3.1.4—The brain as a central information integration organ C3.1.5—The spinal cord as an integrating centre for unconscious processes C3.1.6—Input to the spinal cord and cerebral hemispheres through sensory neurons C3.1.7—Output from the cerebral hemispheres to muscles through motor neurons C3.1.8—Nerves as bundles of nerve fibres of both sensory and motor neurons C3.1.9—Pain reflex arcs as an example of involuntary responses with skeletal muscle as the effector C3.1.10—Role of the cerebellum in coordinating skeletal muscle contraction and balance C3.1.11—Modulation of sleep patterns by melatonin secretion as a part of circadian rhythms	5+2	7	9

			<p>C3.1.12—Epinephrine (adrenaline) secretion by the adrenal glands to prepare the body for vigorous activity</p> <p>C3.1.13—Control of the endocrine system by the hypothalamus and pituitary gland</p> <p>C3.1.14—Feedback control of heart rate following sensory input from baroreceptors and chemoreceptors</p> <p>C3.1.15—Feedback control of ventilation rate following sensory input from chemoreceptors</p> <p>C3.1.16—Control of peristalsis in the digestive system by the central nervous system and enteric nervous system</p> <p>C3.1.17—Observations of tropic responses in seedlings</p> <p>C3.1.18—Positive phototropism as a directional growth response to lateral light in plant shoots</p> <p>C3.1.19—Phytohormones as signalling chemicals controlling growth, development and response to stimuli in plants</p> <p>C3.1.20—Auxin efflux carriers as an example of maintaining concentration gradients of phytohormones</p> <p>C3.1.21—Promotion of cell growth by auxin</p> <p>C3.1.22—Interactions between auxin and cytokinin as a means of regulating root and shoot growth</p> <p>C3.1.23—Positive feedback in fruit ripening and ethylene production</p>			
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		<p>C3.2 Defence against disease</p>	<p>C3.2.1—Pathogens as the cause of infectious diseases  C3.2.2—Skin and mucous membranes as a primary defence  C3.2.3—Sealing of cuts in skin by blood clotting  C3.2.4—Differences between the innate immune system and the adaptive immune system  C3.2.5—Infection control by phagocytes  C3.2.6—Lymphocytes as cells in the adaptive immune system that cooperate to produce antibodies  C3.2.7—Antigens as recognition molecules that trigger antibody production  C3.2.8—Activation of B-lymphocytes by helper T-lymphocytes  C3.2.9—Multiplication of activated B-lymphocytes to form clones of antibody-secreting plasma cells  C3.2.10—Immunity as a consequence of retaining memory cells  C3.2.11—Transmission of HIV in body fluids  C3.2.12—Infection of lymphocytes by HIV with AIDS as a consequence  C3.2.13—Antibiotics as chemicals that block processes occurring in bacteria but not in eukaryotic cells  C3.2.14—Evolution of resistance to several antibiotics in strains of pathogenic bacteria  C3.2.15—Zoonoses as infectious diseases that can transfer from other species to humans  C3.2.16—Vaccines and immunization  C3.2.17—Herd immunity and the prevention of epidemics</p>	5	7	7
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			C3.2.18—Evaluation of data related to the COVID-19 pandemic			
	<b>D. Continuity and change</b>	D3.1Reproduction	<p>D3.1.1—Differences between sexual and asexual reproduction</p> <p>D3.1.2—Role of meiosis and fusion of gametes in the sexual life cycle</p> <p>D3.1.3—Differences between male and female sexes in sexual reproduction</p> <p>D3.1.4—Anatomy of the human male and female reproductive systems</p> <p>D3.1.5—Changes during the ovarian and uterine cycles and their hormonal regulation</p> <p>D3.1.6—Fertilization in humans</p> <p>D3.1.7—Use of hormones in in vitro fertilization (IVF) treatment</p> <p>D3.1.8—Sexual reproduction in flowering plants</p> <p>D3.1.9—Features of an insect-pollinated flower</p> <p>D3.1.10—Methods of promoting cross-pollination</p>	5+3	7	10

			<p>D3.1.11—Self-incompatibility mechanisms to increase genetic variation within a species</p> <p>D3.1.12—Dispersal and germination of seeds</p> <p>D3.1.13—Control of the developmental changes of puberty by gonadotropin-releasing hormone and steroid sex hormones</p> <p>D3.1.14—Spermatogenesis and oogenesis in humans</p> <p>D3.1.15—Mechanisms to prevent polyspermy</p> <p>D3.1.16—Development of a blastocyst and implantation in the endometrium</p> <p>D3.1.17—Pregnancy testing by detection of human chorionic gonadotropin secretion</p> <p>D3.1.18—Role of the placenta in foetal development inside the uterus</p> <p>D3.1.19—Hormonal control of pregnancy and childbirth</p> <p>D3.1.20—Hormone replacement therapy and the risk of coronary heart disease</p>			
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		D3.2Inheritance	<p>D3.2.1—Production of haploid gametes in parents and their fusion to form a diploid zygote as the means of inheritance</p> <p>D3.2.2—Methods for conducting genetic crosses in flowering plants</p> <p>D3.2.3—Genotype as the combination of alleles inherited by an organism</p> <p>D3.2.4—Phenotype as the observable traits of an organism resulting from genotype and environmental factors</p> <p>D3.2.5—Effects of dominant and recessive alleles on phenotype</p> <p>D3.2.6—Phenotypic plasticity as the capacity to develop traits suited to the environment experienced by an organism, by varying patterns of gene expression</p> <p>D3.2.7—Phenylketonuria as an example of a human disease due to a recessive allele</p> <p>D3.2.8—Single-nucleotide polymorphisms and multiple alleles in gene pools</p> <p>D3.2.9—ABO blood groups as an example of multiple alleles</p> <p>D3.2.10—Incomplete dominance and codominance</p>	5+3	7	10
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			<p>D3.2.11—Sex determination in humans and inheritance of genes on sex chromosomes</p> <p>D3.2.12—Haemophilia as an example of a sex-linked genetic disorder</p> <p>D3.2.13—Pedigree charts to deduce patterns of inheritance of genetic disorders</p> <p>D3.2.14—Continuous variation due to polygenic inheritance and/or environmental factors</p> <p>D3.2.15—Box-and-whisker plots to represent data for a continuous variable such as student height</p> <p>D3.2.16—Segregation and independent assortment of unlinked genes in meiosis</p> <p>D3.2.17—Punnett grids for predicting genotypic and phenotypic ratios in dihybrid crosses involving pairs of unlinked autosomal genes</p> <p>D3.2.18—Loci of human genes and their polypeptide products</p> <p>D3.2.19—Autosomal gene linkage</p> <p>D3.2.20—Recombinants in crosses involving two linked or unlinked genes</p> <p>D3.2.21—Use of a chi-squared test on data from dihybrid crosses</p>			
		D3.3Homeostasis	<p>D3.3.1—Homeostasis as maintenance of the internal environment of an organism</p> <p>D3.3.2—Negative feedback loops in homeostasis</p> <p>D3.3.3—Regulation of blood glucose as an example of the role of hormones in homeostasis</p> <p>D3.3.4—Physiological changes that form the basis of type 1 and type 2 diabetes</p> <p>D3.3.5—Thermoregulation as an example of negative feedback control</p>	2+2	3	6

			<p>D3.3.6—Thermoregulation mechanisms in humans</p> <p>D3.3.7—Role of the kidney in osmoregulation and excretion</p> <p>D3.3.8—Role of the glomerulus, Bowman’s capsule and proximal convoluted tubule in excretion</p> <p>D3.3.9—Role of the loop of Henle</p> <p>D3.3.10—Osmoregulation by water reabsorption in the collecting ducts</p> <p>D3.3.11—Changes in blood supply to organs in response to changes in activity</p>			
<b>Practical work</b>				8+8	10	21
<b>4. Ecosystems</b>	<b>A. Unity and Diversity</b>	A4.1 Evolution and speciation	<p>A4.1.1—Evolution as change in the heritable characteristics of a population</p> <p>A4.1.2—Evidence for evolution from base sequences in DNA or RNA and amino acid sequences in proteins</p> <p>A4.1.3—Evidence for evolution from selective breeding of domesticated animals and crop plants</p> <p>A4.1.4—Evidence for evolution from homologous structures</p> <p>A4.1.5—Convergent evolution as the origin of analogous structures</p> <p>A4.1.6—Speciation by splitting of pre-existing species</p> <p>A4.1.7—Roles of reproductive isolation and differential selection in speciation</p> <p>A4.1.8—Differences and similarities between sympatric and allopatric speciation</p> <p>A4.1.9—Adaptive radiation as a source of biodiversity</p> <p>A4.1.10—Barriers to hybridization and sterility of interspecific hybrids as mechanisms for of preventing the mixing of alleles between species</p>	4+1	5	7

			A4.1.11—Abrupt speciation in plants by hybridization and polyploidy			
		A4.2 Conservation of biodiversity	<p>A4.2.1—Biodiversity as the variety of life in all its forms, levels and combinations</p> <p>A4.2.2—Comparisons between current number of species on Earth and past levels of biodiversity</p> <p>A4.2.3—Causes of anthropogenic species extinction</p> <p>A4.2.4—Causes of ecosystem loss</p> <p>A4.2.5—Evidence for a biodiversity crisis</p> <p>A4.2.6—Causes of the current biodiversity crisis</p> <p>A4.2.7—Need for several approaches to conservation of biodiversity</p> <p>A4.2.8—Selection of evolutionarily distinct and globally endangered species for conservation prioritization in the EDGE of Existence programme</p>	3	4	4
	<b>B. Form and function</b>	B4.1 Adaptation to environment	<p>B4.1.1—Habitat as the place in which a community, species, population or organism lives</p> <p>B4.1.2—Adaptations of organisms to the abiotic environment of their habitat</p> <p>B4.1.3—Abiotic variables affecting species distribution</p> <p>B4.1.4—Range of tolerance of a limiting factor</p> <p>B4.1.5—Conditions required for coral reef formation</p> <p>B4.1.6—Abiotic factors as the determinants of terrestrial biome distribution</p> <p>B4.1.7—Biomes as groups of ecosystems with similar communities due to similar abiotic conditions and convergent evolution</p>	3	4	4

			B4.1.8—Adaptations to life in hot deserts and tropical rainforest			
		B4.2 Ecological niches	<p>B4.2.1—Ecological niche as the role of a species in an ecosystem</p> <p>B4.2.2—Differences between organisms that are obligate anaerobes, facultative anaerobes and obligate aerobes</p> <p>B4.2.3—Photosynthesis as the mode of nutrition in plants, algae and several groups of photosynthetic prokaryotes</p> <p>B4.2.4—Holozoic nutrition in animals</p> <p>B4.2.5—Mixotrophic nutrition in some protists</p> <p>B4.2.6—Saprotrophic nutrition in some fungi and bacteria</p> <p>B4.2.7—Diversity of nutrition in archaea</p> <p>B4.2.8—Relationship between dentition and the diet of omnivorous and herbivorous representative members of the family Hominidae</p> <p>B4.2.9—Adaptations of herbivores for feeding on plants and of plants for resisting herbivory</p> <p>B4.2.10—Adaptations of predators for finding, catching and killing prey and of prey animals for resisting predation</p> <p>B4.2.11—Adaptations of plant form for harvesting light</p> <p>B4.2.12—Fundamental and realized niches</p> <p>B4.2.13—Competitive exclusion and the uniqueness of ecological niches</p>	4	5	5
	<b>C. Interaction and interdependence</b>	C4.1 Populations and communities	<p>C4.1.1—Populations as interacting groups of organisms of the same species living in an area</p> <p>C4.1.2—Estimation of population size by random sampling</p>	5	7	7

C4.1.3—Random quadrat sampling to estimate population size for sessile organisms

C4.1.4—Capture–mark–release–recapture and the Lincoln index to estimate population size for motile organisms

C4.1.5—Carrying capacity and competition for limited resources

C4.1.6—Negative feedback control of population size by density-dependent factors

C4.1.7—Population growth curves

C4.1.8—Modelling of the sigmoid population growth curve

C4.1.9—Competition versus cooperation in intraspecific relationships

C4.1.10—A community as all of the interacting organisms in an ecosystem

C4.1.11—Herbivory, predation, interspecific competition, mutualism, parasitism and pathogenicity as categories of interspecific relationship within communities

C4.1.12—Mutualism as an interspecific relationship that benefits both species

C4.1.13—Resource competition between endemic and invasive species

C4.1.14—Tests for interspecific competition

C4.1.15—Use of the chi-squared test for association between two species

C4.1.16—Predator–prey relationships as an example of density-dependent control of animal populations

C4.1.17—Top-down and bottom-up control of populations in communities

C4.1.18—Allelopathy and secretion of antibiotics

		<p>C4.2 Transfers of energy and matter</p>	<p>C4.2.1—Ecosystems as open systems in which both energy and matter can enter and exit  C4.2.2—Sunlight as the principal source of energy that sustains most ecosystems  C4.2.3—Flow of chemical energy through food chains  C4.2.4—Construction of food chains and food webs to represent feeding relationships in a community  C4.2.5—Supply of energy to decomposers as carbon compounds in organic matter coming from dead organisms  C4.2.6—Autotrophs as organisms that use external energy sources to synthesize carbon compounds from simple inorganic substances  C4.2.7—Use of light as the external energy source in photoautotrophs and oxidation reactions as the energy source in chemoautotrophs  C4.2.8—Heterotrophs as organisms that use carbon compounds obtained from other organisms to synthesize the carbon compounds that they require  C4.2.9—Release of energy in both autotrophs and heterotrophs by oxidation of carbon compounds in cell respiration  C4.2.10—Classification of organisms into trophic levels  C4.2.11—Construction of energy pyramids  C4.2.12—Reductions in energy availability at each successive stage in food chains due to large energy losses between trophic levels  C4.2.13—Heat loss to the environment in both autotrophs and heterotrophs due</p>	5	7	7
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			<p>to conversion of chemical energy to heat in cell respiration</p> <p>C4.2.14—Restrictions on the number of trophic levels in ecosystems due to energy losses</p> <p>C4.2.15—Primary production as accumulation of carbon compounds in biomass by autotrophs</p> <p>C4.2.16—Secondary production as accumulation of carbon compounds in biomass by heterotrophs</p> <p>C4.2.17—Constructing carbon cycle diagrams</p> <p>C4.2.18—Ecosystems as carbon sinks and carbon sources</p> <p>C4.2.19—Release of carbon dioxide into the atmosphere during combustion of biomass, peat, coal, oil and natural gas</p> <p>C4.2.20—Analysis of the Keeling Curve in terms of photosynthesis, respiration and combustion</p> <p>C4.2.21—Dependence of aerobic respiration on atmospheric oxygen produced by photosynthesis, and of photosynthesis on atmospheric carbon dioxide produced by respiration</p> <p>C4.2.22—Recycling of all chemical elements required by living organisms in ecosystems</p>			
	<b>D. Continuity and change</b>	D4.1 Natural selection	<p>D4.1.1—Natural selection as the mechanism driving evolutionary change</p> <p>D4.1.2—Roles of mutation and sexual reproduction in generating the variation on which natural selection acts</p> <p>D4.1.3—Overproduction of offspring and competition for resources as factors that promote natural selection</p> <p>D4.1.4—Abiotic factors as selection pressures</p>	2+2	3	6

			<p>D4.1.5—Differences between individuals in adaptation, survival and reproduction as the basis for natural selection</p> <p>D4.1.6—Requirement that traits are heritable for evolutionary change to occur</p> <p>D4.1.7—Sexual selection as a selection pressure in animal species</p> <p>D4.1.8—Modelling of sexual and natural selection based on experimental control of selection pressures</p> <p>D4.1.9—Concept of the gene pool</p> <p>D4.1.10—Allele frequencies of geographically isolated populations</p> <p>D4.1.11—Changes in allele frequency in the gene pool as a consequence of natural selection between individuals according to differences in their heritable traits</p> <p>D4.1.12—Differences between directional, disruptive and stabilizing selection</p> <p>D4.1.13—Hardy–Weinberg equation and calculations of allele or genotype frequencies</p> <p>D4.1.14—Hardy–Weinberg conditions that must be maintained for a population to be in genetic equilibrium</p> <p>D4.1.15—Artificial selection by deliberate choice of traits</p>			
		D4.2Stability and change	<p>D4.2.1—Stability as a property of natural ecosystems</p> <p>D4.2.2—Requirements for stability in ecosystems</p> <p>D4.2.3—Deforestation of Amazon rainforest as an example of a possible tipping point in ecosystem stability</p> <p>D4.2.4—Use of a model to investigate the effect of variables on ecosystem stability</p>	4+2	5	8

			<p>D4.2.5—Role of keystone species in the stability of ecosystems</p> <p>D4.2.6—Assessing sustainability of resource harvesting from natural ecosystems</p> <p>D4.2.7—Factors affecting the sustainability of agriculture</p> <p>D4.2.8—Eutrophication of aquatic and marine ecosystems due to leaching</p> <p>D4.2.9—Biomagnification of pollutants in natural ecosystems</p> <p>D4.2.10—Effects of microplastic and macroplastic pollution of the oceans</p> <p>D4.2.11—Restoration of natural processes in ecosystems by rewilding</p> <p>D4.2.12—Ecological succession and its causes</p> <p>D4.2.13—Changes occurring during primary succession</p> <p>D4.2.14—Cyclical succession in ecosystems</p> <p>D4.2.15—Climax communities and arrested succession</p>			
		D4.3Climate change	<p>D4.3.1—Anthropogenic causes of climate change</p> <p>D4.3.2—Positive feedback cycles in global warming</p> <p>D4.3.3—Change from net carbon accumulation to net loss in boreal forests as an example of a tipping point</p> <p>D4.3.4—Melting of landfast ice and sea ice as examples of polar habitat change</p> <p>D4.3.5—Changes in ocean currents altering the timing and extent of nutrient upwelling</p> <p>D4.3.6—Poleward and upslope range shifts of temperate species</p> <p>D4.3.7—Threats to coral reefs as an example of potential ecosystem collapse</p> <p>D4.3.8—Afforestation, forest regeneration and restoration of peat-</p>	3+1	4	5

			forming wetlands as approaches to carbon sequestration D4.3.9—Phenology as research into the timing of biological events D4.3.10—Disruption to the synchrony of phenological events by climate change D4.3.11—Increases to the number of insect life cycles within a year due to climate change D4.3.12—Evolution as a consequence of climate change			
<b>Scientific Investigation</b>				10	13	13
<b>Total</b>				<b>76 hours SL/ 100 hours HL</b>	<b>98 lessons</b>	<b>129 lessons</b>